

1 **Lipid Extracted Distillers Dried Grains with Solubles (LE-DDGS) as a Partial Replacement**
2 **for Soybean Meal in Hybrid Tilapia (*Oreochromis niloticus* ×**
3 ***Oreochromis aureus*)**

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24

25 **Abstract**

26 Feed costs are primarily driven by the cost of protein sources in the feed. Substitution of
27 expensive protein sources with lower cost ingredients would potentially reduce feed cost. Lipid-
28 extracted distillers dried grains with solubles (LE-DDGS) is a relatively new product of distillers
29 dried grains with solubles (DDGS) which could be used as alternative protein source in tilapia
30 feed formulations. Two growth trials were conducted to evaluate the substitution of soybean
31 meal with LE-DDGS in practical diets for the hybrid tilapia (*O. niloticus* × *O. aureus*). The first
32 trial evaluated five levels of LE-DDGS (0, 20, 30, 40 and 50 g/kg) in a practical diet containing
33 360 g kg⁻¹ protein and 60 g kg⁻¹ lipid. Increasing percentages of LE-DDGS generally resulted in
34 a reduced growth of hybrid tilapia with a significant depression in performance when
35 incorporated in the diet at 40 and 50 g/kg. The second study evaluated high levels of inclusion
36 (0, 20, 40 and 50%) with lysine supplements and a fifth diet with and additional 20 g/kg lipids
37 (2.7 g kg⁻¹ lysine and 80 g kg⁻¹ lipid) with all diets promoting good growth in hybrid tilapia.
38 Overall, results from these studies concluded that LE-DDGS could be a promising protein source
39 in combination with soybean meal in formulated diets containing 360 g kg⁻¹ protein for hybrid
40 tilapia.

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45 **KEY WORDS:** alternative protein sources, nutrient retention, distillers dried grains, ethanol
46 industry

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48 **Introduction**

49 Reducing feed cost in aquaculture is important for the long term sustainability of this
50 industry. One way to reduce costs is to systematically reduce or replace the more expensive
51 components of the feed. This must be done in such a way as to reduce overall production costs
52 while ensuring that such substitution will not compromise the growth performance of fish.
53 Towards this goal, numerous studies have been conducted with the purpose of reducing fish
54 meal-based protein with plant protein sources in feed formulations (Brinker and Reiter, 2011).
55 The use of plant-based proteins in aquatic feeds has increased as they are cost effective protein
56 sources with consistent quality and worldwide availability (Watanabe, 2002). Distillers dried
57 grains with solubles (DDGS) is a co-product from ethanol industry. As the ethanol industries
58 continue to mature, DDGS has been found to be a cost effective protein source in feed
59 formulations. The properties of DDGS offer several potential advantages to animals including
60 moderate protein and lipid contents, as well as phosphorus, vitamins, and trace minerals. Another
61 benefit of DDGS is that it does not contain anti-nutritional factors found in other plant protein
62 sources such as trypsin inhibitor and phytate which are presented in soybean meal (Wilson and
63 Poe, 1985; Shiau et al., 1987) and gossypol which is contained in cottonseed meal (Jauncey and
64 Ross, 1982; Robinson, 1991). That can cause negative impacts to aquatic digestive system and
65 may influence its palatability.

66 Although DDGS contains moderate proportion of protein, it usually contains lower levels
67 of lysine, the most limiting amino acid, compared to fishmeal (Cheng and Hardy, 2004).
68 Therefore, it might be necessary to supplement lysine when using high levels of DDGS as a
69 protein source in fish diets. Numerous research projects have shown that DDGS supplemented

70 with lysine is a suitable protein source in fish diets (Webster et al., 1991; Wu et al., 1996, 1997;
71 Belyea et al., 2004; Lim et al., 2007). The nutrient compositions of the resulting DDGS vary
72 considerably depending on the sources of grains and the production process used in ethanol
73 industry. Corn is the most widely used feedstock for ethanol production in the United States.
74 Other grains such as sorghum (Corredor et al., 2006), wheat (Ojowi et al., 1997; Nyachoti et al.,
75 2005), and barley (Mustafa et al., 2000), are also used in ethanol industry. Due to new and
76 emerging technologies in fuel ethanol production, some plants are modifying their processing to
77 increase the value of ethanol co-products. Extracting the lipids from DDGS (lipid-extracted
78 DDGS, LE-DDGS) allows for a higher protein and amino acid contents. LE-DDGS contains
79 about 440 g kg⁻¹ crude protein and 300 g kg⁻¹ crude fat, while DDGS contains 270 g kg⁻¹ crude
80 protein and 100 g kg⁻¹ crude fat.

81 Information on LE-DDGS as an alternative protein source in aquatic feeds has been
82 limited. Therefore, in this study, two growth trials were conducted to identify the response of
83 hybrid tilapia using corn LE-DDGS with and without lysine supplementation as a partial
84 replacement for soybean meal.

85

86 **Materials and methods**

87 *Experimental diets*

88 In the first trial, six isonitrogenous diets (360 g kg⁻¹ crude protein) were designed to
89 replace soybean meal with corn LE-DDGS at increasing levels of inclusion 0, 200, 300, 400, and
90 500 g kg⁻¹ of diet (D0-1, D200-1, D300-1, D400-1, D500-1) on an isonitrogenous basis (Table
91 1). Lysine was supplemented to diets containing the highest levels of LE-DDGS (D500L-1).
92 Amino acid composition of the experimental diets was reported in Table 2. Corn oil was adjusted

93 to maintain similar lipid level at 60 g kg⁻¹ among dietary treatments. In the second trial, five diets
94 were designed to contain 360 g kg⁻¹ protein and formulated to contain LE-DDGS at increasing
95 levels of 0, 200, 400, and 500 g kg⁻¹ (D0-2, D200-2, D400-2, D500-2) as substitutes for soybean
96 meal (Table 3). Also, lysine was supplemented in diets contained LE-DDGS 400 and 500 g kg⁻¹
97 (D400L-2, D500L-2). Amino acid composition of the experimental diets is reported in Table 4.
98 Corn oil was adjusted to maintain a similar lipid level across all but one diet. In the second trial
99 one diet was formulated to contain 500 g kg⁻¹ LE-DDGS with lysine supplement and an
100 additional 20 g kg⁻¹ lipid. (D500LF-2). All diets were manufactured at the Aquatic Nutrition
101 Laboratory at Auburn University (Auburn, AL, USA) by using standard procedures for the
102 laboratory production of fish diet. Feed ingredients and oil were placed in a food mixer (Hobart
103 Corporation, Troy, OH, USA) for 15 minutes. Hot water was then added to the mixture in order
104 to attain an appropriate consistency for pelleting. Diets went through a 4-mm-diameter meat
105 grinder, dried at 70° C to a moisture content of less than 10%, and stored in the freezer at -20° C
106 until used. Feed samples were collected to determine proximate and amino acids composition
107 following AOAC (1995) procedures.

108

109 ***Experimental fish, feeding, and sampling***

110 Hybrid tilapia (*O. niloticus* × *O. aureus*) were obtained from a commercial fingerling
111 producer in Florida (Aquasafra, Bradenton, FL, USA). Juveniles were reared in nursery tanks
112 before they were acclimated to the control diet for 2 weeks. In the first trial, the experiment
113 consisted of six treatments with four replicates. Juvenile tilapia with average weight 6.00 ± 0.11
114 g (mean ± S.E.M., n=50) were randomly placed into 24 aquaria (50 L) at a stocking density of 25
115 fish per aquarium. Aquaria were supplied with flow-through (0.6–1.0 L min⁻¹), heated, and

116 dechlorinated tap water maintained at a temperature of 27° C. Water was continuously aerated,
117 and photoperiod maintained at 12:12 h light:dark schedule. Water temperature and dissolved
118 oxygen were randomly measured in three random aquaria every other day using an YSI Model
119 58 Oxygen Meter (Yellow Springs Instrument Model 58, Yellow Springs, OH, USA). Fish were
120 fed to apparent satiation twice daily (0800h and 1500h) for an 8-wk period. The amount of feed
121 consumed was recorded daily, calculated by the differences in diet weights prior the first and last
122 feeding. Fish from each aquarium were group weighed and counted to determine weight gain and
123 survival every other week. At the end of the experiment, four fish from each aquarium were
124 randomly sacrificed to determine the whole body composition.

125 In the second trial, juvenile tilapia with average weight 2.23 ± 0.10 g (mean \pm S.E.M.,
126 $n=50$) were placed into 20 tanks (150 L) at a stocking density of 20 fish per tank in a
127 recirculating system. Water temperature and dissolved oxygen were randomly measured twice
128 daily in the morning and afternoon (800h and 1600h) using a YSI Model 58 Oxygen Meter
129 (Yellow Springs Instrument Model 58, Yellow Springs, OH, USA). Fish in four tanks were
130 randomly assigned to each of the five experimental diets. Feed input was calculated between 50
131 to 70 g kg⁻¹ of average fish weight every other week. Test diets were applied twice daily at 0800
132 and 1600 h for a 12-wk experimental period. Fish in each tank were group weighed and counted
133 biweekly to determine weight gain (WG) and survival as well as readjust the daily feed input. At
134 the conclusion of the 12-wk growth trial, fish were counted and group weighed to obtain final
135 weight (FW). Weight gain, feed conversion ratio (FCR) and survival were determined using
136 equation 1.1, 1.2 and 1.3, respectively. Four fish from each aquarium were randomly sampled to
137 determine the whole body composition.

$$138 \quad \text{Weight gain (g)} = \text{Average final weight (g)} - \text{Average initial weight (g)} \quad 1.1$$

$$139 \quad \text{Feed conversion ratio} = \text{Dry feed fed (g)} / \text{Weight gain (g)} \quad 1.2$$

$$140 \quad \text{Survival (\%)} = (\text{Initial fish number} - \text{Final fish number}) \times 100 \quad 1.3$$

141

142 **Analytical Method**

143 Moisture content of both fish and feed samples were determined using equation 2.1, by
 144 recording their original weight (wet), placing them in ceramic crucibles, drying them in
 145 isothermal oven at 105°C for 8 hours, placing them in desiccators until they were at room
 146 temperature and recording their final weight (dry).

$$147 \quad \text{Moisture content (\%)} = \frac{W_w - W_d}{W_w} \times 100 \quad 2.1$$

148 where:

W_w = wet weight of the sample

149 W_d = weight of sample after drying

150

151 Protein content in the first trial was measured by combustion method using an FP-2000
 152 Nitrogen Analyzer (Leco Corporation, St. Joseph, MI, USA). Protein content in the second
 153 experiment was analyzed using The Kjeldahl method (Ma and Zauzago, 1942). Energy contents
 154 of all samples were determined using a bomb calorimeter (1425 Semimicro Calorimeter, Illinois,
 155 USA). Apparent net protein retention (ANPR) and apparent net energy retention (ANER) were
 156 calculated by using equation 2.2 and 2.3, respectively.

157

$$\text{ANPR (\%)} = \frac{(\text{Pf in fish} \times \text{FW} \times \text{DMf of fish}) - (\text{Pi in fish} \times \text{FW} \times \text{DMi of fish})}{\text{P in feed} \times \text{amount of feed consumed}} \quad 2.2$$

158

159 Where:

P_f = Protein in final fish

160 P_i = Protein in initial fish

161 DM_f = Dry matter of final fish (%)

162 DM_i = Dry matter of initial fish (%)

163

$$ANER (\%) = \frac{(E_f \text{ in fish} \times FW \times DM_f \text{ of fish}) - (E_i \text{ in fish} \times FW \times DM_i \text{ of fish})}{E \text{ in feed} \times \text{amount of feed consumed}} \quad 2.3$$

164

165 Where: E_f = Energy in final fish
 166 E_i = Energy in initial fish
 167 DM_f = Dry matter of final fish (%)
 168 DM_i = Dry matter of initial fish (%)

169

170

171 **Statistical Analysis**

172 All data were statistically analyzed using one-way analysis of variance to determine
 173 significant differences ($P < 0.05$) among treatments, which was followed by the Tukey multiple
 174 comparison test to determine significant differences among treatment means. All statistical
 175 analyses were carried out using SAS (V9.2 SAS Institute, Cary, NC, USA).

176

177 **Results**

178 The laboratory trials were conducted without any noticeable problems with water quality.
 179 Water quality parameters were within suitable ranges for the culture of this specie (Table 5). For
 180 the first trial, the growth performance (FW, WG, FCR, survival, ANPR and ANER) were
 181 summarized in Table 6. Fish fed D0-1 had the highest FW and WG, and was significantly higher
 182 than the other treatments ($P < 0.05$), except the D200-1 and D300-1 treatment. Survival ranged
 183 from 98 to 100%. No significant differences were found in survival among all treatments. Feed
 184 conversion ratio ranged from 0.91 to 1.02. The lowest FCR was found in the D0-1 treatment, and
 185 the highest FCR in the D500-1 treatment. In regards to ANPR and ANER, control diet (D0-1)

186 had the highest ANPR and ANER and was significantly higher than other treatments, except the
187 D300-1 treatment for ANPR and the D200-1 treatment for ANER. ANPR and ANER ranged
188 from 39.90 to 46.35 % and 34.81 to 43.02 %, respectively.

189 Growth performance of hybrid tilapia in Trial II was summarized in Table 7. The
190 response of juvenile tilapia fed at increasing levels of LE-DDGS (D0-2, D200-2, D400L-2,
191 D500L-2, and D500LF-2) were not significantly different with regards to FW, WG, FCR,
192 survival, ANPR, and ANER ($P>0.05$). The D500LF-2 treatment had the highest FW, WG, and
193 ANER, though these were not significantly different among treatments. The D500L-2 treatment
194 had the lowest FW, WG, and FCR. The D200-2 treatment had the lowest ANPR and ANER. At
195 the conclusion of the experimental period, FW ranged from 55.82 to 65.55 g. WG ranged from
196 2,469.03 to 2,808.73 %. FCR ranged from 1.04 to 1.14. Survival ranged from 77.10 to 89.75%.
197 ANPR and ANER ranged from 35.60 to 41.93% and 32.06 to 34.25%, respectively.

198

199 Discussion

200 A number of fish feeding studies evaluating the efficacy of DDGS from ethanol industry
201 have been conducted. Several studies have evaluated composition of DDGS in order to estimate
202 nutritional value as a feedstuff (Chevanan et al., 2005; Rosentrater and Muthukumarappan,
203 2006). Belyea et al. (2004) analyzed DDGS samples from 1997 to 2001, and found that protein
204 and lipid levels ranged from 283 to 333 g kg⁻¹ and 109 to 126 g kg⁻¹, respectively. Likewise,
205 Spiehs et al. (2002) evaluated DDGS samples from 1997 to 1999 and concluded an average
206 protein and lipid concentration of 302 g kg⁻¹ and 109 g kg⁻¹, which were higher than those
207 reported in the past at the level of 281 g kg⁻¹ protein and 82 g kg⁻¹ lipid. There is a trend of
208 increasing DDGS protein and lipid resulted from new technology in ethanol production. Due to

209 new developments and technology advancements, some ethanol plants are modifying their
210 processing and extracting lipid from DDGS resulting in an increased protein and reduced lipid
211 contents. Although DDGS has relatively high fiber content, tilapia can utilize carbohydrates
212 more efficiently than cultured piscivorous fishes (NRC, 1983; Lim and Webster, 2006).

213 Distillers dried grains with solubles has been utilized in aquatic feeds since 1940s;
214 however, inclusion levels in diets were fairly low (Phillips, 1949; Phillips et al., 1964). Result
215 from the first trial indicated that diet inclusion rates of 200, 300, 400, and 500 g kg⁻¹ LE-DDGS
216 caused a reduced performance on juvenile tilapia growth. This might be due to the fact that LE-
217 DDGS contains low level of lipid which can provide less energy to the diets than DDGS. Results
218 from several studies demonstrated that the inclusion of DDGS in aquatic feed can be utilized up
219 to 300 g kg⁻¹ of diet. Wu et al. (1994) observed that tilapia fry fed a commercial diet (360 g kg⁻¹
220 crude protein, fish-based protein) had a lower weight gain than fish fed diet containing 290 g kg⁻¹
221 DDGS formulated with 360 g kg⁻¹ crude protein. Coyle et al. (2004) concluded that diet (300 g
222 protein kg⁻¹ of diet) containing DDGS at a level of 300 g kg⁻¹ of diet in combination with meat
223 and bone meal and soybean meal provided an effective performance to hybrid tilapia (*O.*
224 *niloticus* × *O. aureus*). Similarly, Zhou et al. (2010) evaluated fuel-based DDGS to replace
225 soybean meal and corn meal in juvenile hybrid catfish diet (320 g protein kg⁻¹ of diet). They
226 suggested that diet containing 300 g kg⁻¹ DDGS also provided good growth, protein retention,
227 and feed conversion in catfish. Result from the present study also demonstrated that LE-DDGS
228 can be used in hybrid tilapia diet at the inclusion level of 300 g kg⁻¹ without causing any negative
229 effects on fish growth.

230 Results from the first experiment revealed that juvenile tilapia offered a diet with 500 g
231 kg⁻¹ LE-DDGS had significantly lower FW and WG ($P < 0.05$) as compared to fish fed the control

232 diet (D0-1). The supplementation of lysine (D500L-1) did not appear improve final weights of
233 the fish. This indicates that either crystalline lysine was not utilized or there was another more
234 limiting nutrient, possibly energy. Results of the second experiment indicated an effective
235 utilization of all the diets and hence the lysine supplement by tilapia fed high level of LE-DDGS.
236 There were no significant differences between performance of fish fed control diet (D0-2) and
237 diets containing 400 and 500 g kg⁻¹ LE-DDGS with lysine supplementation (D400L-2 and
238 D500L-2). Although not significant there is a general trend of reduced weight gain as LE-DDGS
239 is increased as was seen in the first experiment.

240 **Robinson and Li (2008)** evaluated a combination of DDGS and cotton seed meal as a
241 replacement of soybean meal, and demonstrated that diet (290 g kg⁻¹ crude protein) containing
242 300-400 g kg⁻¹ DDGS with lysine supplement could be used to replace soybean meal without an
243 adverse effect on performance of catfish. **Webster et al. (1991)** fed channel catfish diets with
244 increasing levels of DDGS (0, 350, and 700 g kg⁻¹ of diet), and they found that channel catfish
245 fed a diet with 700 g kg⁻¹ DDGS grew significantly less than other dietary treatments. However,
246 fish fed diet containing 700 g kg⁻¹ DDGS with 4 g kg⁻¹ lysine had a similar growth compared to
247 fish fed diets containing 0 and 350 g kg⁻¹ DDGS. **Shelby et al. (2008)** reported the growth
248 performance of tilapia fed diets (325 g kg⁻¹ crude protein) with increasing levels of DDGS (0,
249 300, and 600 g kg⁻¹) as a replacement of a combination of soy and corn meals. This study
250 showed that diet containing 600 g kg⁻¹ DDGS with no lysine supplementation led to a negative
251 growth performance of tilapia. The addition of lysine to the 600 g kg⁻¹ DDGS diet improved fish
252 weight gain to a level that was not significantly different from control fish.

253 Distillers grains are moderate protein and energy sources, however, the information on
254 energy availability for aquatic animals is often limited. **Smith et al. (1980)** reported digestible

255 energy of distillers dried soluble in rainbow trout was 2,436 kcal kg⁻¹ diet. There is no data on the
256 energy and protein digestibility of DDGS in tilapia; therefore, in these studies, the digestible
257 energy and digestible protein of DDGS was calculated based on the numbers of rainbow trout as
258 reference. Cheng and Hardy (2004) conducted an experiment on the effect of microbial phytase
259 in corn-based DDGS on apparent digestibility coefficients (ADCs) in rainbow trout and found
260 that ADCs of crude fat, crude protein, gross energy, minerals, and amino acids in DDGS
261 supplemented with different dosages of phytase were 78.9-88.9, 80.0-91.9, 50.5-66.6, 7.3-99.7,
262 73.9-96.8%, respectively. Smith et al. (1980) conducted experiment in rainbow trout (*Salmo*
263 *gairdneri*), and reported that their digestibility of corn DGS was 71.9% for protein and 58.6% for
264 carbohydrate (Lovell, 1977). In the first trial, the addition of crystalline lysine to diet containing
265 500 g kg⁻¹ LE-DDGS (D500L-1) did not improve fish growth. Fish fed diet containing 500 g kg⁻¹
266 LE-DDGS with lysine supplement (D500L-1) obtain similar growth performance with fish fed
267 diet containing 500 g kg⁻¹ LE-DDGS without lysine supplement (D500-1), and obtained FW,
268 WG ANPR, and ANER lower than fish fed the basal diet (D0-1). This suggested a limitation in
269 other nutrients which retard growth of fish. DDGS contains relatively high crude fiber, which
270 could result in a limited use as an energy source. An earlier study reported that Nile tilapia fry
271 exhibited the best growth with a protein to energy ration of 110 mg kcal⁻¹ (El-Sayed and Teshima
272 1992). Furthermore, Kubaryk (1980) pointed out that the protein to energy ratios between 108 to
273 120 mg kcal⁻¹ led to the best growth in Nile tilapia fry. Estimating GE and DE indicated that, as
274 inclusion levels of LE-DDGS in the diets increased from 200 to 500 g kg⁻¹ of diets, the ratio of
275 digestible energy to protein generally decreased approximately from 10.77 to 8.42 kcal g⁻¹
276 digestible protein or 111.1 to 133.3 mg digestible protein kcal⁻¹ (Table 8). In the second
277 experiment, the ratio of digestible energy to protein was increased by adding 20 g kg⁻¹ lipid to

278 the diet containing 500 g kg⁻¹ LE-DDGS with lysine from 7.94 to 8.27 kcal g⁻¹ digestible protein
279 (Table 9). Thus, the poor growth of fish fed diets containing increasing level of LE-DDGS
280 observed in this study is likely due to a deficiency in dietary digestible energy. As lipid content
281 in diet contained 500 g kg⁻¹ LE-DDGS with lysine supplement increased from 60 (D500L-2) to
282 80 g kg⁻¹ (D500LF-2), weight gain of juvenile tilapia improved more than 300% compared to
283 fish fed diet contained 500 g kg⁻¹ LE-DDGS with lysine (D500L-2). **Stickney and Wurts (1986)**
284 stated that the performance of *O. aureus* could be improved as fish oil was provided at 75-100 g
285 kg⁻¹ of the diet; however, best growth was achieved with menhaden oil at 100 g kg⁻¹ of the diet.
286 **Chou and Shiau (1996)** observed that 5 g kg⁻¹ of dietary lipid was sufficient to meet the minimal
287 requirement of juvenile hybrid tilapia (*O. aureus* × *O. niloticus*), but a level of 12 g kg⁻¹ was
288 needed for maximal growth. **Jauncey and Ross (1982)** reported that the diet containing lipid in
289 excess of 12 g kg⁻¹ caused a depressed growth of hybrid tilapia (*O. aureus* × *O. niloticus*).

290 Results of the present study, demonstrate that LE-DDGS can be incorporated into
291 practical diets for hybrid tilapia at 300 g kg⁻¹. Furthermore, results indicate that if 500 g kg⁻¹ LE-
292 DDGS are utilized, energy may be limiting and that increasing the digestible energy content of
293 the diet may improve performance.

294

295

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398 **Table 1** Ingredient compositions (g 100 g⁻¹ as is) of seven experimental diets formulated to
 399 contain 360 g kg⁻¹ protein and 60 g kg⁻¹ lipid used in trial I. Diets contained increasing levels of
 400 LE-DDGS (0, 200, 300, 400, and 500 g kg⁻¹) as well as lysine supplement in diets with high
 401 levels of LE-DDGS (500 g kg⁻¹ of diet) as a substitute for soybean meal. Proximate (% as is),
 402 were analyzed at Midwest Laboratories, Inc. (Omaha, NE, USA).

Ingredient	D0-1	D200-1	D300-1	D400-1	D500-1	D500L-1
Menhaden fishmeal ¹	5.00	5.00	5.00	5.00	5.00	5.00
Soybean meal ²	49.80	39.80	34.80	29.80	24.80	24.20
DDGS-lipid extracted ³	0.00	20.00	30.00	40.00	50.00	50.00
Menhaden fish oil ¹	0.82	0.82	0.82	0.82	0.80	0.82
Corn oil	2.92	1.74	1.15	0.56	0.00	0.00
Whole wheat ⁴	33.06	24.74	20.58	16.42	12.25	12.58
Trace mineral premix ⁵	0.50	0.50	0.50	0.50	0.50	0.50
Vitamin premix ⁶	0.80	0.80	0.80	0.80	0.80	0.80
Choline chloride ⁴	0.20	0.20	0.20	0.20	0.20	0.20
Stay C 25% ⁷	0.10	0.10	0.10	0.10	0.10	0.10
Dicalcium phosphate ⁸	2.30	1.80	1.55	1.30	1.05	1.05
Soy lecithin ⁹	0.50	0.50	0.50	0.50	0.50	0.50
Corn gluten meal ¹⁰	4.00	4.00	4.00	4.00	4.00	4.00
L-Lysine HCl ¹¹	0.00	0.00	0.00	0.00	0.00	0.25
Crude protein	35.70	37.80	36.70	37.40	36.60	36.80
Crude fat	6.61	6.86	6.33	6.19	6.57	6.37

Crude fiber	2.69	4.03	3.52	3.79	4.62	2.79
Moisture	8.56	6.37	7.62	5.42	6.70	6.81
Ash	6.69	6.72	6.25	6.40	6.66	6.17

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- 403 ¹ Omega Protein Inc., Reedville, VA, USA.
² Faithway Feed Co., Guntersville, AL, USA.
³ Poet Dakote Gold Inc., Sioux Falls, SD, USA.
⁴ Gold Medal, General Mills Inc., Minneapolis, MN, USA.
⁵ Trace mineral (g/100g Premix): Cobalt chloride 0.004, Cupric sulfate pentahydrate 0.25, Ferrrous sulfate 4.0, Magnesium sulfate anhydrous 13.862, Manganous sulfate monohydrate 0.65, Potassium iodide 0.067, Sodium selenite 0.01, Zinc sulfate heptahydrate 13.193, cellulose 67.964.
⁶ Vitamin (g/kg Premix): Thiamin HCl 0.44, Riboflavin 0.63, Pyridoxine HCl 0.91, D pantothenic acid 1.72, Nicotinic acid 4.58, Biotin 0.21, Folic acid 0.55, Inositol 21.05, Menadione sodium bisulfite 0.89, Vitamin A acetate (500,000 IU g⁻¹) 0.68, Vitamin D₃ (400,000 IU g⁻¹) 0.12, DL-alpha-tocopherol acetate (250 IU g⁻¹) 12.63, cellulose 955.59.
⁷ Stay-C® (L-ascorbyl-2-polyphosphate), **DSM, Pendergrass GA USA**
⁸ MP Biochemicals Inc., Solon, OH, USA.
⁹ Solae Company, St. Louis, MO, USA.
¹⁰ Grain Processing Corporation, Muscatine, IA, USA.
¹¹ Aldrich-Sigma, St. Louis, MO, USA.

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Table 2 Amino acid composition (g 100 g⁻¹ as is) of the experimental diets containing increasing levels of LE-DDGS (0, 200, 300, 400 and 500 g kg⁻¹) as well as lysine supplement in diets with high levels of LE-DDGS (500 g kg⁻¹ of diet).

Component	D0-1	D200-1	D300-1	D400-1	D500-1	D500L-1
Alanine	1.52	1.72	1.88	2.08	2.22	2.07
Arginine	2.39	2.19	2.29	2.29	2.06	2.20
Aspartic Acid	3.40	3.08	2.83	2.93	2.71	2.85
Cystine	0.54	0.79	0.83	0.65	0.65	0.69
Glutamic Acid	6.56	6.53	6.25	6.31	6.15	6.23
Glycine	1.45	1.48	1.49	1.54	1.55	1.50
Histidine	0.90	1.06	0.92	1.11	1.17	1.06
Isoleucine	1.41	1.49	1.45	1.41	1.36	1.37
Leucine	2.69	2.97	3.07	3.23	3.18	3.31
Lysine	1.83	1.82	1.73	1.72	1.61	1.81
Methionine	0.53	0.63	0.66	0.66	0.66	0.66
Phenylalanine	1.61	1.68	1.82	1.72	1.51	1.63
Proline	2.28	2.52	2.57	2.74	2.66	2.73
Serine	1.83	1.75	1.53	1.78	1.59	1.78
Threonine	1.46	1.38	1.28	1.39	1.29	1.37
Tyrosine	1.33	1.35	1.47	1.48	1.31	1.42
Tryptophan	0.31	0.34	0.31	0.27	0.30	0.30

Diets were analyzed by Midwest Laboratories, Inc., Omaha, NE, USA.

406 **Table 3** Ingredient compositions (g 100 g⁻¹ as is) of five experimental diets formulated to contain
 407 360 g kg⁻¹ protein and 60 g kg⁻¹ lipid used in trial II containing increasing levels of LE-DDGS (0,
 408 200, 400 and 500 g kg⁻¹) with lysine supplement in diets with high levels of LE-DDGS (400, and
 409 500 g kg⁻¹ of diet) as well as lipid supplement in diets containing 500 g kg⁻¹ LE-DDGS with
 410 lysine supplement as a substitute for soybean meal. Proximate (% as is), were analyzed at
 411 Midwest Laboratories, Inc. (Omaha, NE, USA).

Ingredient	D0-2	D200-2	D400L-2	D500L-2	D500LF-2
Menhaden fishmeal ¹	5.00	5.00	5.00	5.00	5.00
Soybean meal ²	48.90	38.98	28.72	23.60	24.50
DDGS-lipid extracted ³	0.00	20.00	40.00	50.00	50.00
Menhaden Fish Oil ¹	0.82	0.82	0.82	0.80	0.80
Corn Oil	3.13	1.92	0.70	0.12	2.15
Whole wheat ⁴	33.55	25.18	17.07	12.61	9.73
Trace Mineral premix ⁵	0.50	0.50	0.50	0.50	0.50
Vitamin premix ⁶	0.80	0.80	0.80	0.80	0.80
Choline chloride ⁴	0.20	0.20	0.20	0.20	0.20
Stay C 25% ⁷	0.10	0.10	0.10	0.10	0.10
Dicalcium phosphate ⁸	2.50	2.00	1.45	1.50	1.45
Soy lecithin ⁹	0.50	0.50	0.50	0.50	0.50
Corn Gluten meal ¹⁰	4.00	4.00	4.00	4.00	4.00
L-Lysine HCl ¹¹	0.00	0.00	0.14	0.27	0.27
Crude protein	35.30	37.20	34.50	33.60	34.00

Crude fat	6.90	7.13	7.10	6.79	8.95
Crude fiber	5.42	4.74	5.17	6.99	6.20
Moisture	9.66	9.95	10.14	11.30	11.68
Ash	7.18	7.15	6.71	6.89	6.62

412 ¹ Omega Protein Inc., Reedville, VA, USA.

² Faithway Feed Co., Guntersville, AL, USA.

³ Poet Dakote Gold Inc., Sioux Falls, SD, USA.

⁴ Gold Medal, General Mills Inc., Minneapolis, MN, USA.

⁵ Trace mineral (g/100g Premix): Cobalt chloride 0.004, Cupric sulfate pentahydrate 0.25, Ferric sulfate 4.0, Magnesium sulfate anhydrous 13.862, Manganous sulfate monohydrate 0.65, Potassium iodide 0.067, Sodium selenite 0.01, Zinc sulfate heptahydrate 13.193, cellulose 67.964.

⁶ Vitamin (g/kg Premix): Thiamin HCl 0.44, Riboflavin 0.63, Pyridoxine HCl 0.91, D pantothenic acid 1.72, Nicotinic acid 4.58, Biotin 0.21, Folic acid 0.55, Inositol 21.05, Menadione sodium bisulfite 0.89, Vitamin A acetate (500,000 IU g⁻¹) 0.68, Vitamin D₃ (400,000 IU g⁻¹) 0.12, DL-alpha-tocopherol acetate (250 IU g⁻¹) 12.63, cellulose 955.59.

⁷ Stay-C[®] (L-ascorbyl-2-polyphosphate), Roche Vitamins Inc., Parsippany, NJ, USA.

⁸ MP Biochemicals Inc., Solon, OH, USA.

⁹ Solae Company, St. Louis, MO, USA.

¹⁰ Grain Processing Corporation, Muscatine, IA, USA.

¹¹ Aldrich-Sigma, St. Louis, MO, USA.

Table 4 Nutrient composition (g 100 g⁻¹ as is) of the experimental diets containing increasing levels of LE-DDGS (0, 200, 400 and 500 g kg⁻¹) with lysine supplement in diets with high levels of LE-DDGS (400, and 500 g kg⁻¹ of diet) as well as lipid supplement in diets containing 500 g kg⁻¹ LE-DDGS with lysine supplement.

Component	D0-2	D200-2	D400L-2	D500L-2	D500LF-2
Alanine	1.72	1.90	2.03	2.08	2.06
Arginine	2.27	2.18	1.99	1.87	1.87
Aspartic Acid	3.32	3.27	2.91	2.79	2.71
Cystine	0.51	0.54	0.55	0.55	0.52
Glutamic Acid	6.69	6.39	5.98	5.55	5.38
Glycine	1.61	1.66	1.57	1.52	1.53
Histidine	0.89	0.92	0.92	0.91	0.90
Isoleucine	1.49	1.49	1.44	1.40	1.38
Leucine	2.94	3.15	3.36	3.46	3.39
Lysine	1.93	1.85	1.82	1.83	1.83
Methionine	0.57	0.63	0.62	0.63	0.60
Phenylalanine	1.79	1.82	1.81	1.81	1.79
Proline	2.20	2.33	2.42	2.40	2.36
Serine	1.52	1.54	1.52	1.51	1.49
Threonine	1.31	1.36	1.32	1.34	1.31
Tyrosine	1.19	1.24	1.25	1.26	1.23
Tryptophan	0.44	0.42	0.37	0.31	0.36

Diets were analyzed by Midwest Laboratories, Inc., Omaha, NE, USA.

Table 5 Summary of water quality variables for growth trials with *O. niloticus* reared under flow-through (trial I) and recirculating system (trial II). Values represent the average, standard deviation, minimum, and maximum readings.

Parameter	Average	Standard deviation	Minimum	Maximum
<i>First trial</i>				
<i>(flow-through system)</i>				
DO (mg L ⁻¹) ^a	6.06	0.46	4.50	6.80
Temperature (°C)	27.68	0.73	25.00	29.20
<i>Second trial</i>				
<i>(recirculating system)</i>				
DO (mg L ⁻¹) ^a	5.95	0.56	4.05	6.89
Temperature (°C)	28.21	1.78	23.50	30.80
Salinity (ppt)	0.30	0.45	0.10	2.00
TAN (mg L ⁻¹) ^b	0.26	0.31	0.06	1.19
Nitrite (mg L ⁻¹)	0.41	0.40	0.03	1.35

^aDissolved oxygen.

^bTotal ammonia-nitrogen

Table 6 Response over a 8-wk growth period for hybrid tilapia (6.0 ± 0.11 g, initial weight), fed diets containing levels of LE-DDGS (0, 200, 300, 400, and 500 g kg⁻¹ of diet) as a substitute for soybean meal, reared under a flow-through system in indoor tanks (Trial I).

Diet	FW (g)	WG (%)	Survival (%)	FCR ^a	ANPR ^b	ANER ^b
D0-1	81.38 ^a	1238.15 ^a	100.00	0.91 ^a	46.35 ^a	43.02 ^a
D200-1	76.47 ^{ab}	1157.08 ^{ab}	100.00	0.93 ^a	41.79 ^b	39.54 ^{ab}
D300-1	75.58 ^{ab}	1156.27 ^{ab}	100.00	0.94 ^a	42.61 ^{ab}	37.61 ^b
D400-1	73.50 ^b	1118.30 ^b	98.00	0.97 ^{ab}	41.32 ^b	36.32 ^b
D500-1	72.30 ^b	1094.46 ^b	98.00	1.02 ^b	39.90 ^b	34.81 ^b
D500L-1	72.02 ^b	1125.62 ^b	99.00	0.97 ^{ab}	40.58 ^b	36.12 ^b
P-value ^c	0.0081	0.0058	0.6691	0.0008	0.0015	0.0014
PSE ^d	1.66	22.86	1.22	0.01	0.91	1.17

^aFeed conversion ratio = Total feed offered / biomass increase.

^bApparent net protein retention and apparent net energy retention

^cAnalysis of variance was used to determine significant differences ($P < 0.05$) among treatment means ($n=4$).

^dPooled standard error of treatment means

Table 7 Response over a 12-wk growth period for hybrid tilapia (2.23 ± 0.11 g, initial weight), fed diets containing levels of LE-DDGS (0, 200, 400, and 500 g kg⁻¹ of diet) as a substitute for soybean meal, reared under a closed recirculating system in indoor tanks (Trial II).

Diet	FW (g)	WG (%)	Survival (%)	FCR ^a	ANPR ^b (%)	ANER ^b (%)
D0-2	63.15	2701.26	77.10	1.08	38.03	32.66
D200-2	61.03	2672.92	87.42	1.14	35.60	32.06
D400L-2	61.09	2645.15	89.75	1.08	38.51	33.89
D500L-2	55.82	2469.03	85.00	1.04	41.93	32.73
D500LF-2	65.55	2808.73	78.30	1.05	40.45	34.25
P-value ^c	0.47	0.78	0.40	0.53	0.32	0.66
PSE ^d	1.67	84.69	2.41	0.02	0.96	0.53

^aFeed conversion ratio = Total feed offered / biomass increase.

^bApparent net protein retention and apparent net energy retention

^cAnalysis of variance was used to determine significant differences ($P < 0.05$) among treatment means ($n=4$).

^dPooled standard error of treatment means

Table 8 Estimated energy and protein contents, as well as digestible energy, digestible protein, and ratio of digestible energy to digestible protein in diets containing increasing levels of LE-DDGS (0, 200, 300, 400 and 500 g kg⁻¹).

Diet	GE ^a (kcal/g)	CP ^b (%)	DP ^c (%)	DE ^d (kcal/g)	DE:CP (kcal/g)	DE:DP (kcal/g)
D0-1	4.31	35.70	29.98	3.23	9.05	10.77
D200-1	4.49	37.80	31.05	3.03	8.04	9.78
D300-1	4.67	36.70	31.59	2.94	8.01	9.31
D400-1	4.61	37.40	32.12	2.84	7.61	8.86
D500-1	4.67	36.60	32.66	2.75	7.51	8.42
D500L-1	4.54	36.80	32.43	2.75	7.48	8.49

^aGE = Gross energy

^bCP = Crude protein

^cDP = Digestible protein

^dDE = Digestible energy

Table 9 Estimated energy and protein contents, as well as digestible energy, digestible protein, and ratio of digestible energy to digestible protein in diets containing increasing levels of LE-DDGS (0, 200, 400 and 500).

Diet	GE ^a (kcal/g)	CP ^b (%)	DP ^c (%)	DE ^d (kcal/g)	DE:CP (kcal/g)	DE:DP (kcal/g)
D0-2	4.43	35.70	29.64	3.04	8.52	10.26
D200-2	4.45	37.80	30.74	2.85	7.54	9.26
D400L-2	4.68	36.70	31.72	2.66	7.24	8.38
D500L-2	4.56	37.40	32.18	2.55	6.83	7.94
D500LF-2	4.71	36.60	32.28	2.67	7.30	8.27

^aGE = Gross energy

^bCP = Crude protein

^cDP = Digestible protein

^dDE = Digestible energy

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